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## Association of current smoking with airway inflammation in chronic obstructive pulmonary disease and asymptomatic smokers

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### Abstract

**Background:** Inflammation in the airways and lung parenchyma underlies fixed airway obstruction in chronic obstructive pulmonary disease. The exact role of smoking as promoting factor of inflammation in chronic obstructive pulmonary disease is not clear, partly because studies often do not distinguish between current and ex-smokers.

**Methods:** We investigated airway inflammation in sputum and bronchial biopsies of 34 smokers with chronic obstructive pulmonary disease (9 Global initiative for Chronic Obstructive Lung Disease stage 0, 9 stage I, 10 stage II and 6 stage III) and 26 asymptomatic smokers, and its relationship with past and present smoking habits and airway obstruction.

**Results:** Neutrophil percentage, interleukin-8 and eosinophilic-cationic-protein levels in sputum were higher in chronic obstructive pulmonary disease (stage I-III) than asymptomatic smokers. Inflammatory cell numbers in bronchial biopsies were similar in both groups. Current smoking correlated positively with macrophages: in bronchial biopsies in both groups, and in sputum in chronic obstructive pulmonary disease. Pack-years smoking correlated positively with biopsy macrophages only in chronic obstructive pulmonary disease.

**Conclusion:** Inflammatory effects of current smoking may mask the underlying ongoing inflammatory process pertinent to chronic obstructive pulmonary disease. This may have implications for future studies, which should avoid including mixed populations of smokers and ex-smokers.

### Background

Chronic obstructive pulmonary disease (COPD) is one of the most important causes of death and its prevalence is still increasing [1]. The major risk factor in the develop-

ment and progression of COPD is cigarette smoking. COPD is characterised by fixed airway obstruction and respiratory symptoms, i.e. chronic cough, sputum production and dyspnoea. COPD patients have, just like so called

healthy smokers, an inflammatory reaction involving the entire tracheobronchial tree [2,3].

As compared to healthy non-smokers the degree of airway inflammation seems higher in COPD patients. For example, higher numbers of CD8 positive T-cells, macrophages, neutrophils, and mast cells, both in central and peripheral airways have been found in COPD patients, irrespective whether these patients were current smokers or ex-smokers [4-10]. In addition, the percentage of neutrophils and IL-8 levels in sputum and broncho-alveolar lavage of COPD patients were higher [7,11-16]. As compared to healthy smokers, the differences with COPD patients are less clear cut. For example, higher numbers of neutrophils, macrophages and CD8 positive T-cells in the peripheral airways of COPD patients were found as compared to smokers [10,17-19], whereas others did not [10,19,20]. Two studies showed a higher percentage of neutrophils and higher IL-8 levels in broncho-alveolar lavage of COPD patients [13,21], whereas Linden et al. found no differences [7]. A few studies showed higher numbers of neutrophils [22], CD3, CD4 [23] CD8 positive T-cells [23,24] in bronchial biopsies, whereas other studies found no differences in neutrophils [24], CD3, CD4 [22,24] and CD8 positive T-cells [22], macrophages, eosinophils and mast cells [22,24]. In conclusion, COPD patients have a higher degree of airway inflammation compared to non-smokers, however it remains unclear whether this is also true comparing COPD patients with so called healthy smokers.

Definite conclusions about the exact role of cigarette smoking in COPD are difficult to draw for a number of reasons. First, most studies investigated smokers combined with ex-smokers. Second, many studies investigated COPD patients combined with patients with chronic bronchitis. Third, many studies investigated only one aspect of inflammation, or only one compartment (sputum, broncho-alveolar lavage, bronchial biopsies, peripheral airways), which may be insufficient to obtain a full view. Fourth, remodelling in COPD may itself generate and maintain an inflammatory process, independent of cigarette smoking [25].

In order to elucidate the role of smoking on inflammation in COPD we have investigated airway inflammation in sputum and bronchial biopsies of asymptomatic smokers and smokers with COPD. Furthermore, we assessed whether airway inflammation is related to the number of cigarettes smoked per day, to pack-years smoking and to severity of airway obstruction.

## Methods

### Subjects

Subjects were recruited from the pulmonary outpatient clinic of the Groningen University Hospital and by advertisements in local newspapers. 34 smokers with COPD and 26 smokers without COPD were included according to the ERS criteria [26]. Smokers with COPD had chronic cough and sputum production for at least 3 months for 2 successive years, and an forced expiratory volume in one second ( $FEV_1$ )/ vital capacity (VC)  $\leq 88\%$  of predicted for males and  $\leq 89\%$  of predicted for females. Asymptomatic smokers without COPD had no chronic respiratory symptoms, and  $FEV_1/VC > 88\%$  of predicted for males and  $> 89\%$  of predicted for females and an  $FEV_1 > 85\%$  of predicted. To detect respiratory symptoms to delineate the group of symptomatic smokers without COPD we used the questions about respiratory symptoms and smoking from the Dutch version of the British Medical Research Council's standardised questionnaire [27]. These data were collected by interviewing the participants at the first visit. All participants had to meet the following criteria: age between 45–75 years, minimum of 10 pack-years smoking, actual smoking  $\geq 10$  cigarettes per day, reversibility to salbutamol  $< 9\%$  of the predicted  $FEV_1$ , no use of inhaled or oral corticosteroids in the previous 6 months, no atopy (no positive skin prick test for 10 common aeroallergens and serum total IgE  $< 200$  IU), no respiratory tract infection 1 month prior to the study. After inclusion, subjects were categorized according to the Global Initiative for Chronic Obstructive Lung Disease, GOLD stage 0-IV [28]. GOLD stage 0 (symptomatic smokers): 'at risk' with normal spirometry but chronic symptoms (cough, sputum production); GOLD stages I-IV:  $FEV_1/FVC$  post bronchodilator (post BD)  $< 70\%$  and GOLD stage I:  $FEV_1$  post BD  $\geq 80\%$  predicted; GOLD stage II:  $50\% \leq FEV_1$  post BD  $< 80\%$  predicted; GOLD stage III:  $30\% \leq FEV_1$  post BD  $< 50\%$  predicted and GOLD stage IV:  $30\% \leq FEV_1$  post BD or  $FEV_1 < 50\%$  predicted plus respiratory failure. Current smoking was confirmed by urinary cotinine levels  $> 25$  ng/ml. Before each measurement subjects were asked not to use long or short-acting  $\beta_2$  agonists and/or ipratropium at least 12 hours before the test. The local medical ethics committee approved the study protocol and all subjects gave their written informed consent.

### Study Design

All subjects visited the hospital on 5 separate days, at least one week apart. Lung function tests (flow-volume curves, reversibility, airway conductance), airway hyperresponsiveness (AHR) to methacholine and to adenosine-5'-monophosphate (AMP), and sputum induction (twice) were performed and all subjects underwent bronchoscopy.

### Lung Function

Lung function ( $FEV_1$ ,  $FEV_1/VC$ ) was measured using dry wedge spirometry (Masterscope, Jaeger, Breda, The Netherlands) according to standardized guidelines [29]. Specific airway conductance (sGaw) was measured by body plethysmography (Masterscope, Jaeger, Breda, The Netherlands). Provocation tests were performed with a 2-minute tidal breathing method adapted from Cockcroft and co-workers [30]. After an initial nebulised saline (0.9%) challenge, subjects inhaled doubling concentrations, ranging from 0.038 to 39.2 mg/ml of methacholine-bromide (Sigma Chemical Co. St Louis, MO) and from 0.04 to 320 mg/ml of AMP (Sigma Chemical Co. St Louis, MO) at 5-minute intervals. Tests were terminated when  $FEV_1$  had fallen 20% or more from its baseline value ( $PC_{20}$ ).

### Sputum Induction and Sputum Processing

Sputum was induced by inhalation of hypertonic saline aerosol and processed as described previously [31]. Briefly 15 minutes after salbutamol (400 µg) inhalation, hypertonic saline (3%, 4% and 5% w/v) was nebulised and inhaled for each concentration over a period of 7 minutes. Whole sputum samples were processed within 2 hours after termination of the induction. Two sputum cytospin slides were stained with May-Grünwald-Giemsa for differential cell counts. Counting of 600 non-squamous cells in a blinded way by one technician (B.R.). Sputum samples containing > 80% of squamous cells were excluded from analysis as indication of poor cytospin quality. Interleukin 8 (IL-8) concentration was measured using ELISA (CLB, Amsterdam, the Netherlands) and eosinophil cationic protein (ECP) concentration by a fluorezyme immunoassay (ImmunoCAP ECP, Pharmacia, Uppsala, Sweden).

### Bronchoscopy and biopsy processing

Subjects were not allowed to drink or eat at least 4 hours prior to the bronchoscopy. Smoking was not allowed before the bronchoscopy. On arrival,  $FEV_1$  was measured before and 15 minutes after 400 µg salbutamol. Hereafter subjects gargled with 5 ml of 2% lidocaine and had 2% lidocaine sprayed on the posterior pharynx, dripped onto the vocal cords and into the trachea, with a maximum dose of 3 mg/kg lidocaine. A flexible fiberoptic bronchoscope (Olympus B1 IT10, Olympus Optical, Tokyo, Japan) was introduced and preferably 6 bronchial biopsies were taken from the subcarinae of the right middle or lower lobe using a fenestrated cup forceps (Olympus FB-21C, Olympus Optical Tokyo, Japan) [32]. Biopsies were collected into sterile PBS on ice. Two biopsies were directly embedded in Tissue Tek (Bayer Corporation, Elkhart, Indiana, USA), snap-frozen in liquid isopentane and stored at  $-80^\circ\text{C}$ , 4 biopsies were fixed in 4% paraformaldehyde, processed and embedded in paraffin.

Serial sections were cut from the paraffin biopsies with a thickness of 4 µm and stored at room temperature. Selection of morphological optimal tissue was based on a hematoxylin and eosin stained slide. Tissue slides were deparaffinised with xylene (15 min) and dehydrated before staining. Immunohistochemical staining was performed with monoclonal antibodies against: CD3 (A0452, DAKO, Copenhagen, Denmark) CD4 (CD4-368, Novacastra, UK), CD8 (M7103, DAKO), B cells (CD20 L26, M0755, DAKO), mast cell tryptase (AA1, M7052, DAKO), neutrophil elastase (NP57, M0752, DAKO), macrophages (CD68, M0814, DAKO) and secreted form of eosinophilic cationic protein EG2 (Pharmacia Diagnostics, Sweden). Negative controls were obtained by omission of the primary antibody. Slides were pre-treated with 1 mM EDTA buffer pH = 8 (CD4, CD8), 0.1 mM tris-HCL buffer pH = 9.0 (CD20) in the microwave for 8 or 30 minutes respectively or with 1% protease for 30 minutes at room temperature (CD68, NP57, AA1, EG2). CD3 slides were incubated overnight at  $80^\circ\text{C}$  with tris/HCL buffer pH = 9.0. All stainings were performed in an automated system using the Dako Autostainer (DAKO, Copenhagen, Denmark), except for CD4 that was done manually.

The dilutions used were: CD3 1:100; CD4 1:25; CD8 1:100; CD68 1:50; EG2 1:200; NE 1:200; AA1 1:100; CD20 1:400. As detection system we used labelled streptavidin-biotin (LSAB+, K0690, DAKO, Copenhagen, Denmark) except for CD4 where the Envision system (K5007, DAKO, Copenhagen, Denmark) was used. 3-amino-9-Ethyl Carbazole (AEC) (K3469, DAKO, Copenhagen, Denmark), or Nova Red (SK4800, Vector, USA) for CD4, was used as a chromogen (substrate) giving a reddish-brown reaction product. Hydrogen peroxide was used for blocking endogenous peroxidase and haematoxylin was used as a counterstain. For each antigen, all slides were stained simultaneously.

For each immunohistochemical staining 2 sections of 2 different bronchial biopsies were quantified by computer-assisted image analysis at magnification of 200× (Qwin, Leica Microsystems Imaging Solutions Ltd, Cambridge, UK). Automated image analysis to quantify immunopositivity was performed using the next algorithm: first the intensity of the positive area (cells) was appointed in each biopsy by the observer, followed by the intensity of the total area of the biopsy, based on the red-green-blue (RGB) color model [33,34]. After this, all images of the biopsy were analyzed. Excluded were epithelium, submucosal glands, airway smooth muscle tissue and damaged tissue. Afterwards, the algorithm determined the immuno-positive area and the measured area of the biopsy, leading to the percentage positive area per biopsy. A positive area was at least  $11.8\ \mu\text{m}^2$ , to exclude false positive areas. In this manner, the total positive area and the

total measurable area of the biopsy were quantified and the percentage positive area per biopsy was calculated. The smallest evaluable area per section (after exclusion of epithelium, submucosal glands, airway smooth muscles and damaged tissue) was 0.4 mm<sup>2</sup>. The mean percentage positive area of two biopsies was used. Measurements were performed in a blinded way by 2 observers (B.R. and B.W.).

#### **Data analysis**

Analyses were performed using SPSS for Windows 10.0 (SPSS Inc., Chicago, IL). Values of  $p < 0.05$  were considered statistically significant. Clinical data were expressed in means ( $\pm$  SD) or geometric means (minimum-maximum); inflammatory data were expressed in medians (minimum-maximum). Differences between asymptomatic smokers, symptomatic smokers (GOLD 0) and smokers with COPD (GOLD stage I, II and III) were analysed using the Kruskal-Wallis test, a non-parametric equivalent to one-way ANOVA. Only, when the Kruskal-Wallis test was significant the Mann Whitney U test was used to analyse the differences between the 3 groups. Differences between GOLD stages 0, I, II, and III were analysed using the Kruskal-Wallis test, when this test was significant the Mann Whitney U test was used to analyse the differences between the different GOLD stages.

Correlations between smoking characteristics and lung function parameters were calculated with Pearson correlation test. Correlations between inflammatory cells and mediators in sputum and/or bronchial biopsies and smoking characteristics or lung function parameters were calculated with Spearman's rank correlation test. The subjects with GOLD stages I-III were used to investigate the correlations in COPD patients.

## **Results**

### **Asymptomatic smokers versus smokers with GOLD stage 0-III**

The 34 smokers with COPD were categorised into GOLD stage 0 'symptomatic smokers' (n = 9), GOLD stage I (n = 9), stage II (n = 10) and stage III (n = 6); none of the patients fulfilled the criteria for GOLD stage IV. The clinical characteristics of all subjects are presented in (see Additional file 1). Symptomatic smokers (GOLD stage 0) had significantly decreased lung function and more severe hyperresponsiveness to AMP than asymptomatic smokers had. COPD patients in GOLD stages I-III were older, had significantly more pack-years smoking, lower airway conductance and more severe hyperresponsiveness to AMP and methacholine than asymptomatic and symptomatic (GOLD 0) smokers.

### **Sputum**

Two asymptomatic smokers could not produce sputum. The median (range) percentage non-squamous cells was 94 (75-99)% in COPD patients and 88 (64-99)% in asymptomatic smokers (table 1). Symptomatic smokers (GOLD 0) had higher percentage of sputum neutrophils than asymptomatic smokers. Smokers with COPD (GOLD stage I-III) had higher percentage of neutrophils, IL-8 and ECP levels in sputum than asymptomatic smokers, and higher IL-8 levels in sputum than symptomatic smokers. The percentage of macrophages was lower (table 1). In the separate GOLD stages, GOLD stage II had a higher percentage of sputum neutrophils compared with the asymptomatic smokers (70% and 60% respectively) and higher IL-8 and ECP levels in sputum than GOLD stage 0 and I (21.4 ng/ml versus 8.7 and 8.5 ng/ml respectively, and 291  $\mu$ g/L versus 120 and 99  $\mu$ g/L respectively). GOLD stage III had higher levels of IL-8 than GOLD stage 0 (27.7 ng/ml and 8.7 ng/ml respectively) and lower ECP levels than GOLD stage II (87  $\mu$ g/L versus 291  $\mu$ g/L).

### **Bronchial biopsies**

Bronchial biopsies could not be collected or were of insufficient quality in 2 asymptomatic smokers, in 1 subject GOLD stage 0, and in 6 COPD patients. The percentage positive area of inflammatory cells in bronchial biopsies (CD3, CD4, CD8, CD20, neutrophils, macrophages, eosinophils and mast cells) did not differ between COPD (GOLD I-III), symptomatic smokers (GOLD 0) and asymptomatic smokers (table 2). Only COPD patients with GOLD stage II had a higher percentage positive CD3 area than asymptomatic smokers (1.84 (0.24-9.24) and 0.76 (0.17-2.4) respectively).

### **Correlations of lung function with smoking and airway inflammation**

FEV<sub>1</sub> post BD (% predicted) correlated negatively with the number of pack-years smoking ( $r = -0.51$ ,  $p = 0.03$ ) in COPD, but not significantly with the number of cigarettes smoked per day.

FEV<sub>1</sub> post BD correlated negatively with IL-8 levels in sputum and positively with macrophages in sputum and mast cells in bronchial biopsies of patients with COPD (table 3). The latter correlation was mainly caused by 4 patients with low mast cell positive areas. In asymptomatic smokers, no significant correlations were found between lung function and airway inflammation (table 3).

AHR did not correlate with number of cigarettes smoked per day, number of pack-years smoking or airway inflammation in sputum or bronchial biopsies in both asymptomatic smokers and COPD patients (data partially presented and discussed earlier: Willemse et al, [35]).

**Table 1: Sputum inflammation in smokers with COPD, symptomatic smokers and asymptomatic smokers**

	COPD GOLD I-III	Symptomatic smokers GOLD 0	Asymptomatic smokers
<i>Sputum, n</i>	25	9	24
Volume, ml	4.1 (0.7–14.3)	3.1 (0.3–10.0)	2.3 (0.6–10.8)*
Total cells, 10 <sup>6</sup>	6.7 (1.4–54.5)	4.1 (1.1–15.3)	3.5 (0.2–23)*
Cell conc., 10 <sup>3</sup> /ml	1507 (484–9620)	2134 (534–4146)	1445 (303–4592)
Nonsquamous cells, %	94 (75–99.7)	92 (81–96)	88 (64–99.5)
Eosinophils, 10 <sup>3</sup> /ml	15 (0–106)	20 (0–135)	13 (0–235)
%	1.4 (0–4.0)	1.8 (0–4.1)	0.8 (0–12.6)
Neutrophils, 10 <sup>3</sup> /ml	870 (235–7608)	1575 (434–2558)	661 (164–2856)
%	72.6 (45–89)	66 (39–81)	60.1 (31.5–92.6)* †
Macrophages, 10 <sup>3</sup> /ml	407 (89–2615)	535 (89–2422)	568 (22–1488)
%	25.4 (8.2–52.7)	28.7 (16.6–58.4)	36 (6.5–62.4)*
Lymphocytes, 10 <sup>3</sup> /ml	14 (0–77)	15 (0–62)	11 (1–161)
%	0.8 (0.1–4)	0.8 (0.4–1.6)	0.7 (0.1–3.8)
Epithelial cells, 10 <sup>3</sup> /ml	10 (0–107)	0.4 (0–84)	10 (0–55)
%	0.5 (0–11)	0.5 (0–2.5)	0.8 (0–6.6)
Basophils, 10 <sup>3</sup> /ml	0 (0–4)	0 (0–12)	0 (0–8)
%	0 (0–0.1)	0 (0–0.3)	0 (0–0.3)
IL-8, ng/ml	16.8 (2.1–161) †	8.7 (0.1–25.7)	5.3 (0–25)*
ECP, µg/L	157 (32–2700)	119.8 (13.3–238)	66 (4.7–1282)*

Values expressed in median (range). Abbreviations: cell conc. = cell concentration; IL-8 = interleukin-8; ECP = eosinophilic cationic protein.  
\* p < 0.05 asymptomatic smokers versus total COPD (I-III), Mann-Whitney -U test, † p < 0.05 versus GOLD stage 0, Mann-Whitney-U test.

**Table 2: Inflammation in bronchial biopsies from smokers with COPD and asymptomatic smokers**

	Total COPD GOLD I-III	Symptomatic smokers GOLD 0	Asymptomatic smokers
<i>Biopsies, n</i>	19	8	24
CD3, %positive area	1.05 (0.2–9.24)	0.68 (0.19–1.7)	0.76 (0.17–2.4)
CD4, %positive area	0.041 (0.01–0.57)	0.073 (0–0.18)	0.04 (0–0.15)
CD8, %positive area	0.27 (0.03–2.55)	0.19 (0.02–1.53)	0.33 (0.3–1.25)
CD4/CD8 ratio	0.19 (0.1–4.4)	0.39 (0.04–1.2)	0.18 (0–0.91)
CD20, %positive area	0.003 (0–3.40)	0.003 (0–0.23)	0.005 (0–0.12)
NP57, %positive area	0.025 (0–0.13)	0.05 (0–0.23)	0.021 (0–0.36)
CD68, %positive area	0.035 (0–0.21)	0.056 (0–0.16)	0.041 (0–0.32)
EG2, %positive area	0.021 (0–0.31)	0.049 (0–0.15)	0.063 (0–0.59)
AA1, %positive area	0.15 (0.01–0.91)	0.22 (0–0.41)	0.22 (0.1–1.16)

Values expressed in median (range). Abbreviations: CD20 = B-cell marker, NP57 = neutrophil elastase, CD68 = macrophages, EG2 = eosinophils, AA1 = mast cell tryptase.

### Correlations of current smoking with airway inflammation

The number of cigarettes smoked per day correlated negatively with neutrophils and positively with macrophages in sputum, which was significant in COPD (table 3, figure 1). The number of cigarettes smoked per day correlated positively with macrophages in bronchial biopsies, in both groups (table 3, figure 2). In asymptomatic smokers, the number of cigarettes per day correlated negatively with the number and percentage of eosinophils in sputum. In COPD the number of cigarettes smoked per day

correlated negatively with eosinophil area in bronchial biopsies (table 3).

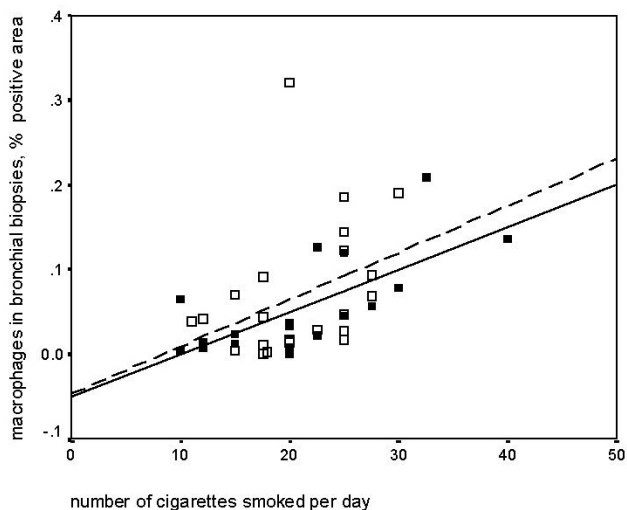
### Correlations of pack-years smoking with airway inflammation

In COPD patients pack-years smoking was positively correlated with the macrophage percentage positive area (table 3). Otherwise no significant correlations were found.

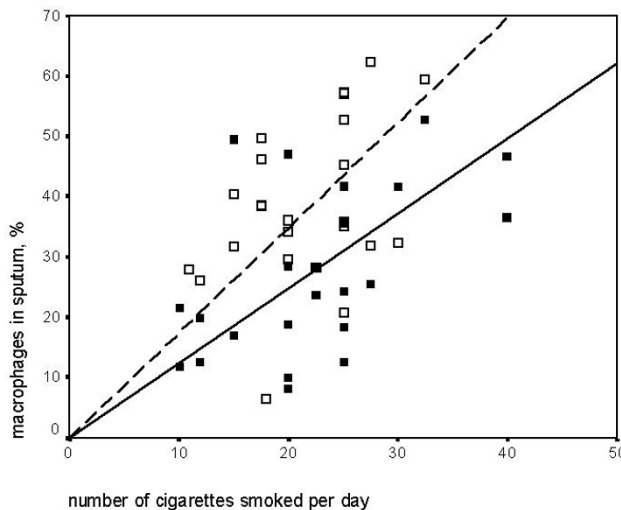
**Table 3: Spearman's rank correlations between current smoking and airway obstruction and airway inflammation.**

	COPD GOLD I-III (n = 19)		Asymptomatic smokers (n = 26)	
	rho	p-value	rho	p-value
<i>Cigarettes/day</i>				
Neutrophils sputum, %	-0.46	0.02	-0.24	NS
Macrophages sputum, %	0.44	0.027	0.35	0.095
Eosinophils sputum, %	-0.19	NS	-0.39	0.057
Eosinophils sputum, 10 <sup>6</sup> /ml	-0.11	NS	-0.44	0.029
CD68 biopsy, % pos. area	0.69	0.002	0.46	0.029
EG2 biopsy, % pos. area	-0.79	0.001	0.02	NS
<i>Pack-years smoking</i>				
CD68 biopsy, %pos. area	0.48	0.043	0.21	NS
<i>FEV<sub>1</sub> post BD, %pred.</i>				
Neutrophils sputum, %	-0.31	NS	0.15	NS
Macrophages sputum, %	0.41	0.046	-0.22	NS
IL-8 sputum, ng/ml	-0.54	0.007	0.15	NS
CD68 biopsy, %pos. area	0.33	NS	0.18	NS
AA1 biopsy, %pos. area	0.53	0.02	0.09	NS

CD68 = macrophages; EG2 = eosinophils; % pos. area = percentage positive area; FEV<sub>1</sub> = forced expiratory volume in one second; post BD = post bronchodilator (15 minutes after 400 µg salbutamol); IL-8 = interleukin 8; AA1 = mast cells; NS = not significant



**Figure 2**  
Spearman's rank correlation: Cigarettes smoked per day and percentage of macrophages in biopsies. COPD (■, —): rho = 0.69 p = 0.002 and asymptomatic smokers(□, - - -): rho = 0.46 p = 0.03.



**Figure 1**  
Spearman's rank correlation: Cigarettes smoked per day and macrophages in induced sputum. COPD (■, —): rho = 0.44 p = 0.03 and asymptomatic smokers(□, - - -): rho = 0.35 p = 0.1.

**Discussion**

This study shows that asymptomatic smokers, symptomatic smokers (GOLD stage 0), and smoking patients with COPD have a large overlap in inflammation as assessed in sputum and airway wall biopsies. Patients

with stage GOLD I-III had a higher percentage of neutrophils, and higher ECP and IL-8 levels in sputum than asymptomatic smokers, and higher IL-8 levels than symptomatic smokers. In symptomatic smokers percentage sputum neutrophils were higher than in asymptomatic smokers.

Whereas current smoking was associated with higher numbers of inflammatory cells in both asymptomatic smokers and COPD patients, pack-years smoking was only associated with higher airway wall macrophages in COPD and to the severity of airway obstruction. More severe airway obstruction in its turn was associated with lower percentage of sputum macrophages in smokers with COPD. Thus, the small difference in airway inflammation found between smokers with and without COPD may be due to the interference of current cigarette smoking.

This study demonstrates that a higher number of daily smoked cigarettes is associated with a higher percentage of macrophages in bronchial biopsies and sputum, both in smokers with COPD and asymptomatic smokers. In addition, eosinophils and neutrophils in sputum were negatively correlated to current smoking. Only few studies have provided data on correlations between airway inflammation and current smoking since smokers and ex-smokers were generally investigated together as one group. Two studies reported a positive correlation between neutrophils in bronchoalveolar lavage and the number of cigarettes smoked per day when asymptomatic smokers, chronic bronchitis patients and COPD patients were analysed together [7,13]. One study in asymptomatic smokers reported that the number of cigarettes smoked per day correlated positively with macrophages and IL-8 levels in bronchoalveolar lavage [36]. Macrophages in the central airways of smokers with and without COPD may be a direct inflammatory reflection of current smoking. On the other hand, it is not likely that current smoking is the only factor responsible for the accumulation of macrophages, since they are also increased in bronchial biopsies of ex-smokers with COPD [37]. Furthermore, we show that not only current smoking but also a higher number of pack-years smoking is associated with higher number of macrophages in COPD. This suggests that effects of current smoking are superimposed upon the underlying macrophage infiltration, which is part of the ongoing inflammatory process in COPD. This is important to realise when investigating the inflammatory and remodelling processes in smokers and ex-smokers with or without COPD. We therefore strongly suggest to avoid including mixed populations of smokers and ex-smokers in future studies on inflammatory processes in COPD

Current smoking was negatively related to eosinophils, i.e. the more cigarettes smoked per day the fewer eosinophils were present in sputum of asymptomatic smokers and in bronchial biopsies of patients with COPD. It may be that smoking has an anti-inflammatory effect on eosinophils or may influence cell kinetics. It has been suggested that carbon monoxide (CO) present in cigarette smoke has an anti-inflammatory potential [38,39], at

least with respect to certain cell types and/or subsets. The extent and relevance of this supposed anti-inflammatory effect in humans remains to be established, but in guinea pigs it has been shown that acute cigarette smoke exposure suppresses the number of eosinophils after 6, 12 and 24 hours [40]. This may indicate that even the cigarettes smoked 24 hours before sputum induction or bronchoscopy could have induced this inverse relationship between current smoking and eosinophilic inflammation, since our participants refrained from smoking for 8 hours before the bronchoscopy. Nevertheless, it is well known that repetitive smoking for several years causes extensive damaging effects, indicating that the long-term overall effects of cigarette smoke dominate the anti-inflammatory effects.

Macrophages in bronchial biopsies of smokers with COPD were positively associated with pack-years smoking. No other relationships between pack-years smoking and airway inflammation were found in our study. This is in agreement with previous studies which either did not find any correlations [41] or did not investigate this [11,15,42]. Only Lams *et al.* [24] reported a positive correlation between CD8+ cells in bronchial biopsies and pack-years smoking and a negative correlation between neutrophils in bronchial biopsies and pack-years smoking, when all smokers (COPD and asymptomatic smokers) were analysed. In broncho-alveolar lavage percentage neutrophils was positively associated with pack-years smoking when all smokers and ex-smokers with and without COPD were analysed together [7,13].

One would expect that in COPD patients inflammatory markers would be more related to pack-years smoking instead of the number of cigarettes smoked per day. However, only macrophages in bronchial biopsies showed a positive correlation with pack-years smoking whereas macrophages, eosinophils and neutrophils were related to the number of cigarettes smoked. This may indicate that some of the inflammation due to cumulative smoke exposure is overruled by inflammation caused by current smoking. Neutrophils and eosinophils are "fast moving, or transient" inflammatory cells, whereas macrophages remain much longer in the lung tissue. This stresses the importance of macrophages in the development and progression of COPD.

This study shows that the percentage of neutrophils in sputum is higher in smokers with COPD (median 72.6%) than in asymptomatic smokers (median 60.1%), especially in GOLD stage II. This is completely in line with results of previous studies, which showed that smokers with moderate to severe COPD had higher total cell numbers and percentages of neutrophils in sputum than asymptomatic smokers [11,15,42]. Thus our finding

suggests that this aspect of inflammation is associated with disease severity.

In symptomatic smokers (GOLD stage 0) the percentage of neutrophils in sputum was higher than in asymptomatic smokers, but similar to COPD patients. This has not been investigated in induced sputum before, however in broncho-alveolar lavage neutrophils show the same pattern [12]. No other differences were found in airway inflammation between symptomatic smokers and asymptomatic smokers. This is in contrast to the study of Sun et al [43], who investigated smokers with chronic bronchitis and found not only an increased number of neutrophils in broncho-alveolar lavage, but also increased eosinophils, mast-cells, CD4 positive and CD8 positive T cells compared to "healthy" smokers. This suggests that chronic bronchitis is better reflected by broncho-alveolar lavage than by induced sputum or bronchial biopsies.

In the present study, IL-8 levels in sputum were significantly higher in smokers with COPD than in asymptomatic and symptomatic smokers. In addition, higher IL-8 levels strongly correlated with more severe airway obstruction in smokers with COPD. This is in line with the data of Keatings *et al.* who showed that both IL-8 and percentage of neutrophils in sputum were increased in patients with moderate COPD as compared to asymptomatic smokers [15]. This may suggest that IL-8, a chemoattractant of neutrophils and an activator of MMP-9, plays a role in the development of airway obstruction. Alternatively, this may reflect the airway obstruction present.

Inflammatory cell density in bronchial biopsies did not significantly differ between smokers with COPD (GOLD I-III), asymptomatic smokers and symptomatic smokers. Only CD3 percentage positive areas in bronchial biopsies were higher in smokers with COPD stage II than in asymptomatic smokers. In agreement with our findings, other studies [24,41] investigating smokers with and without COPD, found no differences in neutrophils, macrophages, eosinophils, CD4 positive cells or CD4/CD8 ratio in bronchial biopsies. In contrast, one previous study demonstrated a higher number of CD8+ cells in smokers with predominantly moderate COPD compared to asymptomatic smokers [24]. In addition, two other studies demonstrated that CD3+ and CD8+ cell numbers were lower and macrophages and neutrophils were higher in smokers with severe COPD [22,41]. It may thus well be that differences between smokers with and without COPD become only apparent in case of severe COPD. Unfortunately the number of patients with evaluable biopsies was too small in our study population (n = 4) to investigate whether this indeed is the case.

A factor that should be taken into consideration is the age difference between the COPD patients and asymptomatic smokers under study. Previous studies investigated younger (mean age 35 years) asymptomatic smokers than our participants (mean age 50 years) [11,15,42]. The composition of sputum may differ between older and younger healthy subjects, as shown in bronchoalveolar lavage where the number of total cells and neutrophils increase with age [44]. Since we investigated COPD patients and asymptomatic smokers of almost similar age, our data are not hampered by age differences.

### Conclusion

Smoking COPD patients with GOLD stage I-III had almost similar airway wall and sputum inflammation as asymptomatic and symptomatic smokers without airway obstruction. Current smoking was associated with airway inflammation in patients with COPD and in asymptomatic smokers, whereas this was not the case for the cumulative pack-years smoked. In contrast, cumulative pack-years smoking was associated with the level of airway obstruction in COPD, suggesting that cumulative smoking induces chronic inflammation with subsequent sequelae of airway obstruction. Our results indicate that inflammatory effects of current smoking may mask findings of chronic inflammation in COPD, since numbers of inflammatory cells in bronchial biopsies and sputum are comparable in smokers with mild COPD and asymptomatic smokers.

### Authors' contributions

BW carried out the data collection and its coordination, immunohistochemical staining and quantification of the bronchial biopsies, performed the statistical analysis and interpretation of the data and drafted and revised the manuscript. NtH contributed to the conception and design of the study, the data collection and the interpretation of the data and revised the manuscript. BR carried out the sputum processing and immunoassays and revised the manuscript. DP contributed to the conception and design of the study, the data collection and the interpretation of the data and revised the manuscript. WT contributed to the conception and design of the study, the data collection and the interpretation of the data and revised the manuscript. All authors read and approved the final manuscript.

### Additional material

#### Additional File 1

Description of the clinical characteristics of the participating subjects

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## References

- Murray CJ, Lopez AD: **Alternative projections of mortality and disability by cause 1990-2020: Global Burden of Disease Study.** *Lancet* 1997, **349**:1498-1504.
- Barnes PJ, Shapiro SD, Pauwels RA: **Chronic obstructive pulmonary disease: molecular and cellular mechanisms.** *Eur Respir J* 2003, **22**:672-688.
- Saetta M, Turato G, Maestrelli P, Mapp CE, Fabbri LM: **Cellular and Structural Bases of Chronic Obstructive Pulmonary Disease.** *Am J Respir Crit Care Med* 2001, **163**:1304-1309.
- Saetta M, Di Stefano A, Maestrelli P, Ferraresso A, Drigo R, Potena A, Ciaccia A, Fabbri LM: **Activated T-lymphocytes and macrophages in bronchial mucosa of subjects with chronic bronchitis.** *Am Rev Respir Dis* 1993, **147**:301-306.
- O'Shaughnessy TC, Ansari TW, Barnes NC, Jeffery PK: **Inflammation in bronchial biopsies of subjects with chronic bronchitis: inverse relationship of CD8+ T lymphocytes with FEV1.** *Am J Respir Crit Care Med* 1997, **155**:852-857.
- Turato G, Di Stefano A, Maestrelli P, Mapp CE, Ruggieri MP, Roggeri A, Fabbri LM, Saetta M: **Effect of smoking cessation on airway inflammation in chronic bronchitis.** *Am J Respir Crit Care Med* 1995, **152**:1262-1267.
- Linden M, Rasmussen JB, Piitulainen E, Tunek A, Larson M, Tegner H, Venge P, Laitinen LA, Brattsand R: **Airway inflammation in smokers with nonobstructive and obstructive chronic bronchitis.** *Am Rev Respir Dis* 1993, **148**:1226-1232.
- Saetta M, Di Stefano A, Turato G, Facchini FM, Corbino L, Mapp CE, Maestrelli P, Ciaccia A, Fabbri LM: **CD8+ T-lymphocytes in peripheral airways of smokers with chronic obstructive pulmonary disease.** *Am J Respir Crit Care Med* 1998, **157**:822-826.
- Pesci A, Rossi GA, Bertorelli G, Aufiero A, Zanon P, Olivieri D: **Mast cells in the airway lumen and bronchial mucosa of patients with chronic bronchitis.** *Am J Respir Crit Care Med* 1994, **149**:1311-1316.
- Saetta M, Turato G, Baraldo S, Zanin A, Braccioni F, Mapp CE, Maestrelli P, Cavalleco G, Papi A, Fabbri LM: **Goblet cell hyperplasia and epithelial inflammation in peripheral airways of smokers with both symptoms of chronic bronchitis and chronic airflow limitation.** *Am J Respir Crit Care Med* 2000, **161**:1016-1021.
- Balzano G, Stefanelli F, Iorio C, De Felice A, Melillo EM, Martucci M, Melillo G: **Eosinophilic inflammation in stable chronic obstructive pulmonary disease. Relationship with neutrophils and airway function.** *Am J Respir Crit Care Med* 1999, **160**:1486-1492.
- Lacoste JY, Bousquet J, Chanez P, Van Vyve T, Simony-Lafontaine J, Lequeu N, Vic P, Enander I, Godard P, Michel FB: **Eosinophilic and neutrophilic inflammation in asthma, chronic bronchitis, and chronic obstructive pulmonary disease.** *J Allergy Clin Immunol* 1993, **92**:537-548.
- Thompson AB, Daughton D, Robbins RA, Ghafouri MA, Oehlkerking M, Rennard SI: **Intraluminal airway inflammation in chronic bronchitis. Characterization and correlation with clinical parameters.** *Am Rev Respir Dis* 1989, **140**:1527-1537.
- Traves SL, Culpitt SV, Russell RE, Barnes PJ, Donnelly LE: **Increased levels of the chemokines GRO $\alpha$  and MCP-1 in sputum samples from patients with COPD.** *Thorax* 2002, **57**:590-595.
- Keatings VM, Collins PD, Scott DM, Barnes PJ: **Differences in interleukin-8 and tumor necrosis factor- $\alpha$  in induced sputum from patients with chronic obstructive pulmonary disease or asthma.** *Am J Respir Crit Care Med* 1996, **153**:530-534.
- Keatings VM, Barnes PJ: **Granulocyte activation markers in induced sputum: comparison between chronic obstructive pulmonary disease, asthma, and normal subjects.** *Am J Respir Crit Care Med* 1997, **155**:449-453.
- Baraldo S, Turato G, Badin C, Bazzan E, Beghe B, Zuin R, Calabrese F, Casoni G, Maestrelli P, Papi A, Fabbri LM, Saetta M: **Neutrophilic infiltration within the airway smooth muscle in patients with COPD.** *Thorax* 2004, **59**:308-312.
- Saetta M, Turato G, Facchini FM, Corbino L, Lucchini RE, Casoni G, Maestrelli P, Mapp CE, Ciaccia A, Fabbri LM: **Inflammatory cells in the bronchial glands of smokers with chronic bronchitis.** *Am J Respir Crit Care Med* 1997, **156**:1633-1639.
- Saetta M, Baraldo S, Corbino L, Turato G, Braccioni F, Rea F, Cavalleco G, Tropeano G, Mapp CE, Maestrelli P, Ciaccia A, Fabbri LM: **CD8+ve cells in the lungs of smokers with chronic obstructive pulmonary disease.** *Am J Respir Crit Care Med* 1999, **160**:711-717.
- de Boer WJ, Sont JK, van Schadewijk A, Stolk J, van Krieken JH, Hiemstra PS: **Monocyte chemoattractant protein 1, interleukin 8, and chronic airways inflammation in COPD.** *J Pathol* 2000, **190**:619-626.
- Pesci A, Balbi B, Majori M, Cacciani G, Bertacco S, Alciato P, Donner CF: **Inflammatory cells and mediators in bronchial lavage of patients with chronic obstructive pulmonary disease.** *Eur Respir J* 1998, **12**:380-386.
- Di Stefano A, Capelli A, Lusuadi M, Balbo P, Vecchio C, Maestrelli P, Mapp CE, Fabbri LM, Donner CF, Saetta M: **Severity of airflow limitation is associated with severity of airway inflammation in smokers.** *Am J Respir Crit Care Med* 1998, **158**:1277-1285.
- Fournier M, Lebargy F, Le Roy LF, Lenormand E, Pariente R: **Intraepithelial T-lymphocyte subsets in the airways of normal subjects and of patients with chronic bronchitis.** *Am Rev Respir Dis* 1989, **140**:737-742.
- Lams BE, Sousa AR, Rees PJ, Lee TH: **Subepithelial immunopathology of the large airways in smokers with and without chronic obstructive pulmonary disease.** *Eur Respir J* 2000, **15**:512-516.
- Rutgers SR, Timens W, Kauffman HF, Postma DS: **Markers of active airway inflammation and remodelling in chronic obstructive pulmonary disease.** *Clin Exp Allergy* 2001, **31**:193-205.
- Stafakas NM, Vermeire P, Pride NB, Paoletti P, Gibson J, Howard P, Yernault JC, Decramer M, Higenbottam T, Postma DS, et al: **Optimal assessment and management of chronic obstructive pulmonary disease (COPD). The European Respiratory Society Task Force [see comments].** *Eur Respir J* 1995, **8**:1398-1420.
- van der Lende R, Orie NG: **The MRC-ECCS questionnaire on respiratory symptoms (use in epidemiology).** *Scand J Respir Dis* 1972, **53**:218-226.
- Global initiative for Chronic Obstructive Lung Disease NIH: **Definitions chapter 1 in Global Initiative for Chronic Obstructive Lung Disease. GOLD 2003:5-10.**
- Quanjer PH, Tammeling GJ, Cotes JE, Pedersen OF, Peslin R, Yernault JC: **Lung volumes and forced ventilatory flows. Report Working Party Standardization of Lung Function Tests, European Community for Steel and Coal. Official Statement of the European Respiratory Society.** *Eur Respir J Suppl* 1993, **16**:5-40.
- Cockcroft DW, Killian DN, Mellon JJ, Hargreave FE: **Bronchial reactivity to inhaled histamine: a method and clinical survey.** *Clin Allergy* 1977, **7**:235-243.
- Rutgers SR, Timens W, Kaufmann HF, van der Mark TW, Koeter GH, Postma DS: **Comparison of induced sputum with bronchial wash, bronchoalveolar lavage and bronchial biopsies in COPD.** *Eur Respir J* 2000, **15**:109-115.
- Aleva RM, Kraan J, Smith M, ten Hacken NH, Postma DS, Timens W: **Techniques in human airway inflammation: quantity and morphology of bronchial biopsy specimens taken by forceps of three sizes.** *Chest* 1998, **113**:182-185.
- Sont JK, de Boer WJ, van Schadewijk WA, Grunberg K, van Krieken JH, Hiemstra PS, Sterk PJ: **Fully automated assessment of inflammatory cell counts and cytokine expression in bronchial tissue.** *Am J Respir Crit Care Med* 2003, **167**:1496-1503.
- Puddicombe SM, Polosa R, Richter A, Krishna MT, Howarth PH, Holgate ST, Davies DE: **Involvement of the epidermal growth factor receptor in epithelial repair in asthma.** *FASEB J* 2000, **14**:1362-1374.
- Willemsse BWM, ten Hacken NHT, Rutgers B, Lesman-Leegte IGAT, Timens W, Postma DS: **Smoking cessation improves direct and indirect airway hyperresponsiveness in COPD.** *Eur Respir J* 2004, **24**:1-6.
- Kuschner WG, D'Alessandro A, Wong H, Blanc PD: **Dose-dependent cigarette smoking-related inflammatory responses in healthy adults.** *Eur Respir J* 1996, **9**:1989-1994.
- Rutgers SR, Postma DS, ten Hacken NH, Kauffman HF, Der Mark TW, Koeter GH, Timens W: **Ongoing airway inflammation in**

- patients with COPD who do not currently smoke. *Thorax* 2000, **55**:12-18.
38. Chapman JT, Otterbein LE, Elias JA, Choi AM: **Carbon monoxide attenuates aeroallergen-induced inflammation in mice.** *Am J Physiol Lung Cell Mol Physiol* 2001, **281**:L209-L216.
  39. Melgert BN, Postma DS, Geerlings M, Luinge MA, Klok PA, Van Der Strate BW, Kerstjens HA, Timens W, Hylkema MN: **Short-term smoke exposure attenuates ovalbumin-induced airway inflammation in allergic mice.** *Am J Respir Cell Mol Biol* 2004, **30**:880-885.
  40. Hulbert WC, Walker DC, Jackson A, Hogg JC: **Airway permeability to horseradish peroxidase in guinea pigs: the repair phase after injury by cigarette smoke.** *Am Rev Respir Dis* 1981, **123**:320-326.
  41. Di Stefano A, Capelli A, Lusuardi M, Caramori G, Balbo P, Ioli F, Sacco S, Gnemmi I, Brun P, Adcock IM, Balbi B, Barnes PJ, Chung KF, Donner CF: **Decreased T lymphocyte infiltration in bronchial biopsies of subjects with severe chronic obstructive pulmonary disease.** *Clin Exp Allergy* 2001, **31**:893-902.
  42. Takanashi S, Hasegawa Y, Kanehira Y, Yamamoto K, Fujimoto K, Satoh K, Okamura K: **Interleukin-10 level in sputum is reduced in bronchial asthma, COPD and in smokers.** *Eur Respir J* 1999, **14**:309-314.
  43. Sun G, Stacey MA, Vittori E, Marini M, Bellini A, Kleimberg J, Mattoli S: **Cellular and molecular characteristics of inflammation in chronic bronchitis.** *Eur J Clin Invest* 1998, **28**:364-372.
  44. Meyer KC, Ershler W, Rosenthal NS, Lu XG, Peterson K: **Immune dysregulation in the aging human lung.** *Am J Respir Crit Care Med* 1996, **153**:1072-1079.

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